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**Low levels of strigolactones in roots as a component of the systemic signal of drought stress in tomato**

Ivan Visentin<sup>a</sup>, Marco Vitali<sup>a</sup>, Manuela Ferrero<sup>a</sup>, Yanxia Zhang<sup>b, c</sup>, Carolien Ruyter-Spira<sup>b</sup>, Ondřej Novák<sup>d</sup>, Miroslav Strnad<sup>d</sup>, Claudio Lovisolo<sup>a</sup>, Andrea Schubert<sup>a</sup>, Francesca Cardinale<sup>a1</sup>

**AUTHOR AFFILIATIONS & INFO**

<sup>a</sup> Laboratory of Plant Physiology, DISAFA - Turin University, 10095 Grugliasco (TO) Italy

<sup>b</sup> Laboratory of Plant Physiology, Wageningen University, 6708 PB Wageningen, The Netherlands

<sup>c</sup> Laboratory of Growth Regulators, Institute of Experimental Botany ASCR & Palacký University Olomouc, Czech Republic

<sup>1</sup>To whom correspondence should be addressed. E-mail: [francesca.cardinale@unito.it](mailto:francesca.cardinale@unito.it); phone +390116708875

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**SHORT TITLE:** Strigolactones in systemic drought signalling

21 **SUMMARY**

- 22 • Strigolactones (SL) contribute to drought acclimatization in shoots, since SL-depleted plants  
23 are hypersensitive to drought due to stomatal hyposensitivity to abscisic acid (ABA).  
24 However, under drought, SL biosynthesis is repressed in roots, suggesting organ specificity  
25 in their metabolism and role. Since SL can be transported acropetally, such drop may also  
26 affect shoots, as a systemic indication of stress.
- 27 • We investigated this hypothesis by analysing molecularly and physiologically WT tomato  
28 scions grafted onto SL-depleted rootstocks, compared to self-grafted WT and SL-depleted  
29 genotypes, during a drought time-course.
- 30 • Shoots receiving few SL from the roots behaved as under mild stress even if irrigated. Their  
31 stomata were hypersensitive to ABA (likely via a localized enhancement of SL synthesis in  
32 shoots). Exogenous SL also enhanced stomata sensitivity to ABA.
- 33 • As the partial shift of SL synthesis from roots to shoots mimics what happens under  
34 drought, a reduction of root-produced SL might represent a systemic signal unlinked from  
35 shootward ABA translocation and sufficient to prime the plant for better stress avoidance.

36

37 **KEYWORDS:** Absciscic acid, Drought, Strigolactones, Systemic signalling, Tomato

38

39

## 40 INTRODUCTION

41 Drought stress counts among the most recurrent and limiting environmental conditions for plant  
42 development and full productivity; under water scarcity, phytohormones cooperatively interact to  
43 allow resource optimization (Christmann *et al.*, 2006). Absciscic acid (ABA) biosynthesis is strongly  
44 and rapidly increased by drought, and prevents water loss mainly by driving stomata closure, thus  
45 controlling transpiration. Also, root-synthesized ABA is, in some plants, a systemic stress signal,  
46 travelling shootward to prevent, among others effects, the negative consequences of soil water  
47 deficit (Comstock, 2002). However, in plants such as *Arabidopsis thaliana* and tomato (*Solanum*  
48 *lycopersicum* L.), ABA produced by roots under water deprivation is unnecessary for shoot  
49 responses, leaving uncertainty on the chemical nature of the systemic drought stress signal  
50 (Holbrook *et al.*, 2002; Christmann *et al.*, 2007). Additionally, it was shown in tomato that ABA  
51 travels from shoots to roots under long-term drought, thus inverting the original hypothesis  
52 (Manzi *et al.*, 2015). Other signals, such as hydraulic, electrical and chemical signals, including  
53 other phytohormones and changes in xylem sap pH, are therefore also thought to contribute  
54 [reviewed by (Huber & Bauerle, 2016)]. It is argued however that positive chemical signals alone  
55 cannot account for the initial stomatal responses to root drying, because of the relatively low  
56 xylem transport velocity (Huber & Bauerle, 2016).

57 Recently, the hormones strigolactones (SL) have been also proposed as signal mediators under  
58 environmental stress. SL have pervasive roles in development, from germination and reproduction  
59 to root and shoot architecture; at various levels, they also promote the interaction with beneficial  
60 root symbionts as well as with detrimental (micro)organisms [reviewed by (Ruyter-Spira *et al.*,  
61 2013)]. SL and ABA share their biosynthetic precursor, both being carotenoid-derived terpenoid  
62 lactones (Matusova *et al.*, 2005). Several enzymes act sequentially in SL biosynthesis: DWARF 27  
63 (D27) is a  $\beta$ -carotene isomerase, CCD7 and CCD8 are Carotenoid-Cleavage Dioxygenases (CCD) and  
64 MORE AXILLARY GROWTH 1 (MAX1) is a class III cytochrome P450 that, with its orthologues and  
65 paralogues and the recently characterized LATERAL BRANCHING OXIDOREDUCTASE (LBO) (Brewer  
66 *et al.*, 2016), is thought to contribute to the oxidation of the SL precursor carlactone and to the  
67 chemical diversification of SL family members [reviewed by (Al-Babili & Bouwmeester, 2015)]. The  
68 core enzyme set is mostly active in roots; root-produced SL are then exported out of the producing  
69 cell by ABC<sub>G</sub> transporter protein(s) such as PhPDR1 (Kretschmar *et al.*, 2012; Sasse *et al.*, 2015),  
70 both to be exuded in soil and to travel shootward, as shown in *Arabidopsis* and tomato (Kohlen *et al.*,  
71 2011). Although transcripts of SL-related genes, and final metabolites, are mostly not or barely

72 detectable in shoots, biosynthesis in above-ground tissues is known to occur, possibly at specific  
73 spots. In fact, wild-type (WT) shoots grafted onto SL-depleted rootstocks do not display the typical  
74 morphological phenotype of SL-depleted plants (Foo *et al.*, 2001; Sorefan *et al.*, 2003).

75 Recently, SL metabolism and physiological effects in plants under osmotic stress conditions have  
76 been analysed. SL-depleted *A. thaliana* and *Lotus japonicus* (Liu *et al.*, 2013) are hypersensitive to  
77 drought at the shoot level, a feature linked to the hyposensitivity of their stomata to endogenous  
78 and exogenous ABA. This finding supports a positive role for SL in the acclimatization to drought in  
79 above-ground organs (Ha *et al.*, 2014; Liu *et al.*, 2015). Consistent with this idea, the transcript of  
80 SL biosynthetic genes is increased by drought in Arabidopsis leaves (Ha *et al.*, 2014). However,  
81 transcription of biosynthetic and SL transporter-encoding genes is repressed along with the  
82 accumulation of SL in non-mycorrhizal *L. japonicus* and tomato roots under drought (Liu *et al.*,  
83 2015; Ruiz-Lozano *et al.*, 2016). This is surprising *per se*, since roots are the main SL production site  
84 under normal conditions; and suggests different dynamics for shoot- and root-derived SL. A  
85 negative correlation between ABA and SL levels was observed in non-mycorrhizal, water-stressed  
86 roots of *L. japonicus* and tomato (Liu *et al.*, 2015; Ruiz-Lozano *et al.*, 2016). Since drought stress-  
87 triggered ABA accumulation is hampered by exogenous SL in *L. japonicus* roots, the drop in SL  
88 biosynthesis in roots under drought might have the role to allow an increase of local ABA and  
89 possibly, also of its levels in the xylem sap, leading to systemic responses to a dropping root water  
90 potential in plants that rely also on ABA for chemical signalling of drought (Liu *et al.*, 2015).  
91 However, the possibility exists that such drop has also a direct physiological effect on shoots,  
92 namely as a systemic indication of stress at the root level, since root-produced SL can also be  
93 transported to the whole plant (Kohlen *et al.*, 2011). This, and the fact that SL are needed locally in  
94 stressed shoots for efficient control of water loss by transpiration (Ha *et al.*, 2014; Liu *et al.*, 2015),  
95 led us to hypothesize that WT scions grafted onto SL-depleted rootstocks may behave as if  
96 stressed even in the absence of stress, at least under some respects, and perform differently  
97 under stress than if grafted onto WT rootstocks.

98 In this work, we investigated the possible systemic significance of the SL decrease in roots under  
99 drought, by analysing molecularly and physiologically WT scions grafted over SL-depleted (CCD7-  
100 silenced) tomato rootstocks, compared to self-grafted WT and SL-depleted genotypes, both under  
101 normal and stress conditions. The results proved that indeed stomata of shoots receiving less SL  
102 from the roots are hypersensitive to ABA also in the absence of stress, possibly through an  
103 enhancement of local SL synthesis. This is likely to mimic what normally happens under drought,

104 and suggests that root-derived SL - or better, a reduction thereof - might be a component of the  
105 systemic signal of stress in tomato.

106

## 107 **MATERIALS AND METHODS**

### 108 ***Plant material and growth conditions***

109 The tomato (*Solanum lycopersicum* L.) *SlCCD7*-silenced line 6936, hereafter called SL-, and its WT  
110 genotype M82 were a kind gift by Dr. H. J. Klee (University of Florida). Seeds were sterilized in 4%  
111 (v:v) sodium hypochlorite containing 0.02% (v:v) Tween 20, rinsed thoroughly with sterile water,  
112 and then germinated for 48 h on moisten filter paper at 25°C in darkness. Subsequently, seedlings  
113 were grown in inert substrate (sand:vermiculite; 1:1, v:v) and the pots watered with Hoagland  
114 solution twice per week. The three grafted lines were produced by the clamp grafting technique  
115 on plants at the 2/4-leaf stage and with stem diameter of about 1.5-2 mm. Water stress was  
116 applied to plants four weeks after grafting by withholding water starting at day zero (T0); shoots  
117 and roots were collected 0, 1, 3 and 5 days after the beginning of the stress (T0 through T5,  
118 respectively; 3 plants per line and sampling point) and stored to -80°C. At T5, 3 plants per line  
119 were watered and collected after 2 additional days to give the rehydrated (recovery) samples. The  
120 experiment was repeated twice. Supporting Information Fig. S1 shows how relative water content  
121 and soil water potential were dropping during the course of one drought experiment. Relative soil  
122 water content was gravimetrically determined by collecting daily ~10 ml of soil from three  
123 different points and depths in each pot (at 5, 10 and 15-cm depth with 120° of angular separation  
124 between each of the respective sample points). The soil was weighed, oven-dried at 100°C for 24 h  
125 and then re-weighed to assess water content. At the same time, the soil water retention curve was  
126 assessed with pressure plate measurements of the potting substrate according to (Tramontini *et*  
127 *al.*, 2014).

128

### 129 ***Gene transcript quantification***

130 Total RNA from tomato roots and shoots was extracted as described (Gambino *et al.*, 2008) and  
131 treated with DNase I (ThermoScientific) at 37°C for 30 min to remove residual genomic DNA. First-  
132 strand cDNA was synthesized from 3 µg of purified total RNA using the High-Capacity cDNA  
133 Reverse Transcription Kit (Applied Biosystems) according to the manufacturer's instructions. For  
134 transcript quantification of *SlCCD7*, *SlCCD8* and *SlNCED1* by quantitative reverse-transcription PCR  
135 (qRT-PCR), the StepOne system (Applied Biosystems) was used, with transcript of the Elongation

factor 1 $\alpha$  (*SIEF1 $\alpha$*  gene) as a reference; primers used are reported as Supporting information in Supplementary Table S1. Three independent biological replicates were analysed and each qRT-PCR reaction was run in technical triplicates. Transcripts of the target genes were quantified by the 2<sup>- $\Delta\Delta C_t$</sup>  method.

### ***Physiological measurements***

Leaf water potential, stomatal conductance and net carbon assimilation were measured daily between 10:00 and 12:00 am on at least three plants per grafted line and independent experiment, as reported by Liu *et al.* (2015). Briefly, stomatal conductance and net carbon assimilation rate were measured with a portable gas exchange system (GFS-3000, Walz GmbH, Effeltrich, Germany) by clamping the most apical leaves of a shoot in the leaf chamber, where photosynthetically active radiation (1200  $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ ), air flow (750  $\mu\text{mol s}^{-1}$ ) and temperature (25°C) were kept constant. Environmental conditions of CO<sub>2</sub> (450 ppm) and vapour pressure deficit (2.3 kPa) were stable during the 10-day experiments. Leaf water potential was measured with a pressure chamber (Scholander *et al.*, 1965) on one leaf per plant, immediately after gas exchange quantification. For the quantification of responses to ABA, stomatal conductance was measured as above at 30-s intervals before and during ABA treatment. This was accomplished by cutting leafy twigs while submerged in filtered water (one leaf each, from three plants per grafted line, treatment and experiment), by letting stomatal conductance stabilize with the twig dipped in water and then by adding ABA to 5, 20 or 50  $\mu\text{M}$  final concentration, while continuously recording every 30 s both stomatal conductance and transpiration rates as detailed above. For treatment with exogenous SL, WT plants were sprayed with a 5  $\mu\text{M}$  solution of *racGR24* (StrigoLab SrL, Turin, IT) 24 h before treatment with ABA 5  $\mu\text{M}$  and stomatal conductance recording as above.

### ***Extraction and quantification of SL and ABA***

Solanacol, orobanchol and didehydro-orobanchol were quantified in the roots of the three grafted lines, while ABA was quantified in both roots and shoots. For SL extractions, 3 plants per line and time-point were pooled, while two independent biological assays were run. For SL quantification, samples (0.5 g each) were manually ground in liquid nitrogen and extracted with 2 ml of cold ethyl acetate containing D6-*epi*-5 deoxystrigol as internal standard (0.05 nmol ml<sup>-1</sup>) in 10-ml glass vials. Standards for didehydro-orobanchol isomers were not available, so quantities for this SL were



expressed as percentage ratio with respect to WT root tissues in the absence of stress (T0); the isomer reported in Fig. S2C is the one with retention time of 4' 6'' in our conditions. The extraction and quantification procedures for SL were performed as previously reported (Lopez-Raez *et al.*, 2010). For ABA extraction, 2 biological replicates of 2 pooled plants each were sampled per line and time-point, while two independent biological assays were run. For ABA quantification, labelled internal standard was added ( $[^2\text{H}]_6$ -ABA, 20 pmol) to each sample (20–25 mg homogenized in 1 ml of cold 10% MeOH in H<sub>2</sub>O, v/v) and subsequently extracted and analysed as detailed (Flokova *et al.*, 2014).

176

## 177 RESULTS

### 178 ***WT shoots transpire and dehydrate less when grafted onto SL-depleted roots***

179 In order to investigate the systemic meaning of SL decrease in stressed roots, we sought to reproduce such condition in the absence of stress. To this purpose, rootstocks of the SL-depleted line SL- (6936) (Vogel *et al.*, 2010) were joined to shoots of the corresponding WT (M82) to give WT/SL- hetero-grafts. Two sets of control plants were also generated, i.e. self-grafts of SL- and WT rootstocks and scions (SL-/SL- and WT/WT, respectively). The physiological, transcriptional and metabolic responses to water stress were examined at different time points for these three sets of individuals. As a preliminary check, SL content in roots was quantified, confirming that the 6936 genotype was indeed defective in SL production (about 20-fold less orobanchol, solanacol and one of the dihydro-orobanchol isomers under unstressed conditions). The three SL metabolites decreased under stress, already one day after water withdrawal, both in WT and SL- roots, irrespectively of the scion genotype (Supporting Information Fig. S2a-c), confirming what observed in PEG-treated *L. japonicus* roots (Liu *et al.*, 2015).

191 Measuring stomatal conductance and leaf water potential confirmed that in tomato, as in Arabidopsis and Lotus, whole-plant SL depletion increases stomatal conductance and decreases leaf water potential in the absence of stress; under the same conditions, WT/SL- plants showed instead significantly lower stomatal conductance than WT/WT (Fig. 1a and T0 in Supporting Information Fig. S3a). Accordingly, leaf water potential values were significantly less negative in WT leaves grafted onto SL- than WT roots (Supporting Information Fig. S3b). Photosynthesis of WT scions grafted over SL- rootstocks was only slightly and non-significantly affected by the reduced gas exchange of hetero-grafts compared to self-grafted WT plants, while both displayed significantly lower values than SL- shoots (Fig. 1b and Supporting Information Fig. S3c).

200 Under stress, the three grafted lines followed a similar trend of stomatal conductance and net  
201 carbon assimilation decrease, although starting from different values (Fig. 1a, b). Under severe  
202 stress, gas exchange in leaves of WT/SL- plants was comparable to the WT, even if leaf water  
203 potential was less negative than in the latter; WT/SL- leaves also performed photosynthesis  
204 significantly better than WT/WT (Supporting Information Fig. S3a-c). SL-/SL- plants confirmed their  
205 hypersensitivity to drought for all parameters tested. These data indicated that SL depletion at the  
206 root level reduces stomatal conductance and attenuates the drop in leaf water potential in WT  
207 shoots under drought, whereas SL depletion in shoots has opposite effects. After rehydration  
208 (Recovery, full symbols in Fig. 1a-b, R in Supporting Information Fig. S3a-c), the physiological  
209 parameters of all three lines returned to levels similar to those observed in the absence of stress.

210

211 ***Both drought and depletion of SL in the roots induce transcript accumulation for SL biosynthetic***  
212 ***genes in the shoots***

213 To assess whether the change in metabolite abundance is regulated at the gene transcription  
214 level, two SL biosynthetic genes (*SICCD7* and *SICCD8*) were profiled by qRT-PCR in roots and shoots  
215 of the three grafted lines under irrigated and drought stress conditions, in the same plant material  
216 used for SL quantification.

217 The analysis confirmed that in roots, transcript amount of both genes inversely correlated with  
218 stress severity for all grafted lines (Fig. 2a, b and Supporting Information Fig. S4a, b). In the shoots  
219 of the same sets of plants however, transcripts of both biosynthetic genes followed an opposite  
220 trend compared to roots and accumulated under drought, as reported previously in *Arabidopsis*  
221 and postulated in *Lotus* (Ha *et al.*, 2014; Liu *et al.*, 2015) (Fig. 2c, d and Supporting Information Fig.  
222 S4d, e). It must be noted however that in terms of relative transcript abundance, values in shoots  
223 remained much lower (about one hundredth; not obvious in the normalized data of Fig. 2) of root  
224 values at T0, even in samples collected under very severe stress at T5. This justifies the fact that  
225 we were unable to detect the final metabolites in these shoot samples (data not shown).  
226 Relevantly here, expression of both biosynthetic genes in WT shoots was significantly higher when  
227 the mutant was used as rootstock (WT/WT vs WT/SL-, Fig. 2c, d and Supporting Information Fig.  
228 S4c, d). This is a known pattern (Johnson *et al.*, 2006) consistent with the idea of a general  
229 negative feedback by the final metabolites on the SL biosynthetic pathway and supported by the  
230 repressive effect of exogenous SL on the same genes [see for example (Liu *et al.*, 2015)]. Overall,  
231 data on transcript of SL-biosynthetic genes indicated that the response of shoots to SL deficiency

in roots overlaps with the response to osmotic stress. In fact, both drought stress and depletion of SL in the roots in the absence of stress induced transcript accumulation of SL biosynthetic genes in tomato shoots.

As an additional observation, *SICCD7* transcripts in unstressed SL- (*CCD7*-silenced) rootstocks were more abundant in grafts bearing a WT instead of a SL- shoot (WT/SL- vs SL-/SL-; T0 of Fig. 2a). This correlated with a very slight increase of SL metabolites, especially orobanchol (see T0, Fig. S2a-c) and suggested that a SL-dependent, shoot-to-root signal feeding back on the transcription/transcript stability of this gene exists in tomato as in Arabidopsis and pea (Foo *et al.*, 2005; Johnson *et al.*, 2006), where it was shown to depend on the *RMS2* locus. Also, *SICCD8* transcripts were more abundant in SL- than WT roots (as expected, given the already mentioned negative feedback of SL on the transcription of their biosynthetic genes; Fig. 2b); and in SL- roots, *SICCD8* transcripts were more concentrated in the presence of a SL- than of a WT scion (Fig. 2b). In this sense, expression of *SICCD7* and *SICCD8* in the root seemed influenced oppositely by the ability of the shoot to produce SL. We may hypothesize that not only locally-produced, but also shoot-synthesized SL may participate (directly or indirectly) in the negative feedback on *SICCD8* expression in the root, and thus that in SL- roots, the presence of a WT scion may lead to less pronounced overexpression of *SICCD8* than in the presence of a SL- scion. Finally, it is noteworthy that the concentration of *SICCD8* transcript in WT shoots grafted onto SL- roots was as high as in SL- shoots in the absence of stress (T0, Fig. 2d) but remained stable along the time-course in the former while it was further induced in the latter (Supporting Information Fig. S4d). We have no easy explanation for this pattern, which might however be due to the fact that leaves of WT/SL- plants dehydrate less and produce less ABA (see further on) along the time-course, than either self-grafted control line.

### ***The low-transpiration phenotype of hetero-grafted, WT/SL- plants is not due to increased total free ABA***

To determine whether the effects of SL depletion on WT shoots may be due to altered ABA metabolism, we set to quantify this hormone in roots and shoots of plants in the three grafted sets. Previous data in Arabidopsis and tomato leaves, and in Lotus roots and shoots, indicated no changes or slight decreases of ABA correlated with SL depletion in shoots, especially under stress (Ha *et al.*, 2014; Liu *et al.*, 2015); ABA content was reported to be lower than in WT under non-stressful conditions only in *CCD8*-silenced tomato shoots (Torres-Vera *et al.*, 2014).

Results showed that under normal conditions, WT roots contain less free ABA than SL- ones (WT/WT vs SL-/SL- and WT/SL- plants, T0 in Supporting Information Fig. S5a) per gram of fresh tissue weight. As stress increased, ABA started accumulating in roots of SL-/SL- and WT/WT plants more quickly than in roots of WT/SL- plants, where ABA was significantly less concentrated than in the roots of the other grafts (Supporting Information Fig. S5a). Correlation curves to leaf water potential values were however substantially superimposable (Fig. 3a). Transcript quantification for *SINCE1*, a key biosynthetic gene for stress-induced ABA in tomato (Munoz-Espinoza *et al.*, 2015), showed good correlation with free ABA content but for a few points and grafting combinations (Fig. 3b and Supporting Information Fig. S5b). These discrepancies between *SINCE1* transcript amounts and ABA concentration may be due to post-transcriptional regulation of biosynthetic enzymes, and/or to the activity of catabolic genes, for example, or to the release/sequestration of free ABA from/in conjugated forms [reviewed by (Xiong & Zhu, 2003)].

While in the absence of stress SL- shoots contained more ABA per gram of fresh weight than WT ones, as stress proceeded and leaf water potential started becoming more negative ABA levels increased faster in WT than in SL- scions; at the moment of maximum stress, ABA concentration was minimum in WT scions grafted onto SL- rootstocks, and intermediate in SL- shoots (Fig. 3c and T5 in Supporting Information S5c). The same trend is seen for transcripts of *SINCE1*, which again showed a good correlation with free ABA content but for a few points and grafting combinations (Fig. 3d and Supporting Information Fig. S5d). These results confirmed that especially under stress, SL depletion in the shoot partially compromises the ability to synthesize ABA. Furthermore, coupled to the physiological data in Fig. 1, they strongly suggested that the low-gas exchange phenotype of hetero-grafted WT/SL- plants was not due to increased free ABA content, given the comparatively low ABA concentration in their tissues.

#### ***WT scions are hypersensitive to ABA if grafted onto SL-depleted rootstocks***

To explore whether altered sensitivity to ABA might rather underlie the physiological and metabolic results described above, shoot sensitivity to exogenous ABA in dependence of the rate of SL production in the roots was investigated. ABA at different concentrations was applied to and absorbed by excised petioles of composite leaves of the three grafted lines, while measuring the time required for the stomata to start closing. This assay on the one hand confirmed in tomato what was already known in *Arabidopsis* and *Lotus*, i.e. that SL-depleted scions are hyposensitive to ABA (at all three - but more convincingly at the lower - concentrations tested), with respect to WT

(SL-/SL- vs WT/WT; Fig. 4). On the other hand, the same analysis proved also that WT scions are indeed hypersensitive to ABA if grafted onto SL- instead of WT rootstocks (WT/SL- vs WT/WT, Fig. 4), as hypothesized on the basis of the stomatal conductance and shoot ABA quantification experiments reported above (Fig. 1a and 3c vs Supporting Information S3a and S5c). We also tested (at 5  $\mu$ M ABA, the concentration for which differences among our lines were more evident) if a pre-treatment with the synthetic SL analogue *rac*GR24 could by itself increase sensitivity to ABA, in a complementary way to SL depletion decreasing it. This was indeed the case (WT/WT plants, GR24-treated vs untreated, Fig. 4).

These data confirmed that the physiological phenotype displayed by the WT/SL- plants both under irrigated and drought conditions was more likely due to a higher sensitivity to endogenous ABA, rather than to its absolute levels. This effect could be linked to a local increase of SL synthesis, given the higher transcript concentration for SL biosynthetic genes under these conditions, and – as a more indirect indication - the fact that ABA sensitivity increased in stomata treated with exogenous SL.

## DISCUSSION

### ***Low SL in the roots prime shoots for drought stress avoidance in tomato***

In this study, we investigated in tomato the possible systemic implications of the drop in SL synthesis happening in roots under osmotic stress. A parsimonious starting hypothesis was that SL depletion in roots could directly or indirectly act as a signal of stress for the shoots. On this basis, hetero-grafted plants with WT scions and SL-depleted rootstocks were to behave as at least mildly stressed, even in the absence of stress. Our physiological data are in agreement with this theory: stomatal conductance values of WT shoots grafted onto SL-depleted rootstocks are significantly lower than those of WT shoots self-grafted onto WT rootstocks in irrigated conditions, and are accompanied by less negative leaf water potential values and, as expected, higher intrinsic water use efficiency (defined as the ratio between net carbon assimilation and stomatal conductance; Supporting Information Fig. S3d). These data support the idea that SL depletion in root tissues affects (directly or indirectly) the physiological response in the shoot and leading to better acclimatization to drought. The ability of shoots to produce SL is needed for this to happen, as stomatal conductance is increased instead when the whole plant (and not only the roots) are *CCD7*-silenced; indeed, this latter condition rather leads to drought hypersensitivity, as shown in

327 SL-depleted Arabidopsis, Lotus and now, tomato plants [(Ha *et al.*, 2014; Liu *et al.*, 2015); this  
328 work].

329

### 330 ***Low SL in the roots and (high) SL in the shoot render stomata hypersensitive to ABA***

331 To determine whether the effects of root SL depletion on WT shoots may be due to altered ABA  
332 levels, this hormone was quantified in roots and shoots of plants in the three grafted sets. SL-  
333 depleted roots and especially shoots contain significantly more ABA per gram of fresh weight than  
334 the WT ones in the absence of stress. Our results in unstressed shoots are in apparent  
335 contradiction to the ones reported on *CCD8*-silenced tomato plants, where shoots of SL-depleted  
336 lines had lower ABA content (Torres-Vera *et al.*, 2014); the most likely explanation is that our data  
337 were normalized over fresh and not dry weight as in Torres-Vera *et al.* In any case during severe  
338 stress, free ABA increases less in tissues of self-grafted SL- than WT plants, a trend already  
339 observed in Lotus (Liu *et al.*, 2015); such situation, coupled to the hyposensitivity to the hormone,  
340 will certainly exacerbate the drought sensitivity of SL-depleted shoots. Instead, the slower and less  
341 pronounced ABA increase in roots and shoots of WT/SL- plants compared to the other lines is in  
342 agreement with the physiological conditions of these plants (which being primed for better stress  
343 resilience, perform better thus needing less ABA). It is of course possible that ABA levels in guard  
344 cells may not be reflected by the total levels of free ABA in the whole leaf tissue, given the strong  
345 compartmentalization of the hormone in different cell types and compartments (Hartung & Slovik,  
346 1991); and thus, that WT/SL- plants had lower  $g_s$  because of locally enhanced ABA accumulation.  
347 However, the results of the ABA-feeding experiment rather supported the hypothesis that such  
348 phenotype was (at least partly) due to stomatal hypersensitivity to the hormone. Finally, the same  
349 experiments also highlighted that SL in the shoot are not only needed but also sufficient to  
350 increase stomatal sensitivity to ABA.

351

### 352 ***Hormonal cross-talk and systemic signalling under drought: fitting SL in the picture***

353 Since our experimental set-up mimics what normally happens during drought, we propose that  
354 these findings are relevant to stress resistance, at least in plants such as Lotus and tomato, for  
355 which a drop in SL synthesis is recorded in roots experiencing osmotic stress or drought. Such drop  
356 might promote a pre-alerted (primed) status in the shoots, which become more sensitive to ABA  
357 at the guard cell level. This message may be conveyed directly (see below) or indirectly, i.e.  
358 through a second messenger that ought to be, at least in tomato, different than ABA. It is to be

359 noted here that SL were proven to cross-talk with other hormones, such as auxins, cytokinins,  
360 brassinosteroids and ethylene, in processes different than drought responses and stomatal closure  
361 (Cheng *et al.*, 2013); and that each of these hormones was shown to affect stomatal aperture  
362 locally (Daszkowska-Golec & Szarejko, 2013). Root-synthesized cytokinins were even proposed to  
363 act as a systemic signal promoting stomatal opening, in a similar way to SL (Davies & Zhang, 1991);  
364 however SL- mutants display reduced cytokinin levels in the shoot, which is the opposite of what  
365 one would expect from a mediator of SL effect (because cytokinins promote stomata aperture,  
366 and SL- shoots transpire more than WT) (Foo *et al.*, 2007). Additionally, shoots were proven to  
367 possess powerful homeostatic mechanisms for the regulation of cytokinin levels, that are largely  
368 unlinked from their concentration in xylem sap (Foo *et al.*, 2007). Resuming, we cannot exclude  
369 that the effect of SL on stomatal closure may be at least partly indirect, i.e. mediated by any of  
370 these hormones, or by other signals yet (and indeed, sensitivity to ABA does play a role). It would  
371 be indeed interesting to quantify other hormones in leaves of our lines, or even better to visualize  
372 their activity in guard cells; and to measure whether, for example, the xylem sap pH in hetero-  
373 grafted plants is different than in self-grafted [possibly, more basic as in droughted tomato plants  
374 (Wilkinson *et al.*, 1998)]. It remains clear that plant hormones, if capable of travelling over long  
375 distances, have a slow propagation velocity in comparison with hydraulic and/or electrical signals.  
376 However, the fact itself that in our model, stomatal closure is rather induced by the lack of an  
377 inhibitor in the shootward flow is attracting, because its decrease might be perceived faster than  
378 flow speed would predict for a positive modulator. In fact, the flow is slowed down by drought,  
379 thus adding to the decrease of the inhibitor itself; additionally, given that SL are degraded upon  
380 perception (Hamiaux *et al.*, 2012), they should be quickly depleted locally unless *de novo* synthesis  
381 or translocation occurs. Finally, expression pattern and intracellular location of the SL  
382 transporter(s) might add another regulation level, for mobility through living tissues.

383 As regards the activity of SL biosynthetic genes, shoots of irrigated, hetero-grafted WT/SL- plants  
384 behave as if under drought, i.e. show increased transcripts of *CCD7* and *CCD8*. These increases in  
385 gene activity might be due to the relief of direct repression of SL synthesis in the shoots by  
386 translocated, root-synthesized SL; a known pattern [e.g. (Johnson *et al.*, 2006; Liu *et al.*, 2013)],  
387 which might itself trigger SL accumulation at specific spots in the shoot (undetectable in whole-  
388 tissue analyses). Even if it is at present impossible to overcome the technical limitations that make  
389 the quantification of SL unfeasible in shoots, we propose that hypersensitivity to ABA in stomata  
390 of WT/SL- plants might be causally linked to higher production of SL in (limited tissue zones of) the

shoot, since i) transcription of SL-biosynthetic genes is activated in WT shoots during stress, but also under non-stressful conditions if WT shoots are grafted onto SL- rootstocks; ii); sensitivity to ABA converts from higher to lower than normal, if not only roots but also shoots are SL-depleted, proving that SL synthesis in the shoots is needed for the effects on ABA sensitivity; iii) exogenous GR24 treatment is sufficient to induce stomatal hypersensitivity to ABA. This latter effect is opposite to the one caused by genetically-due SL depletion, and would explain GR24 ability to confer drought resistance in WT Arabidopsis (Ha *et al.*, 2014). The importance of SL produced in the shoot has been proposed also in branching, because micro-grafting of WT Arabidopsis scions on SL-defective rootstocks does not lead to an increased branching phenotype, as expected if SL synthesis is compromised in the whole plant (Foo *et al.*, 2001; Sorefan *et al.*, 2003). Whether osmotic/drought stress in the absence of such decrease in root-synthesized SL is able to stimulate a similar shoot response, is still to be determined. A schematic drawing of our model is represented in Fig. 5. This model obviously implies that the shoot is able to discriminate between root- and shoot-produced SL; this ability needs to be proven experimentally, but could rely on differential loading in the upstream flow, and/or organ-specific production of the structurally different SL molecules, which make up species-specific SL blends and whose ecological and physiological meanings remain largely unexplored (Kohlen *et al.*, 2011; Kohlen *et al.*, 2012; Bharti *et al.*, 2015; Brewer *et al.*, 2016). Alternatively, or in parallel, the uneven/non-overlapping distribution of the receptor protein D14 and/or of SL transporter(s) in the plant might account for discrimination between locally and distally produced SL (Chevalier *et al.*, 2014; Sasse *et al.*, 2015). From a practical point of view, it remains to be assessed how such graft combinations will perform under other or combined stress. It is important to note on this regard that they will undoubtedly be advantageous in soil infested by parasitic weeds; that not all SL-depleted genotypes are also significantly compromised in mycorrhization (a possible detrimental side-effect); and that with respect to SL synthesis, drought overrules P deficiency under combined stress (Kohlen *et al.*, 2012; Liu *et al.*, 2015). Nonetheless, our results highlight once more the importance of rootstocks in influencing shoot traits, and how they could be exploited to improve crop performances under stress (Albacete *et al.*, 2015; Cantero-Navarro *et al.*, 2016).

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## AUTHOR CONTRIBUTION

F.C. conceived of the work and designed research supported by C.L. and A.S.; I.V. performed research helped by M.V. and M.F.; Y.Z. and O.N. analysed data; C.R.-S. and M.S. provided logistic support to metabolite analyses; I.V. and F.C. wrote the paper. All authors read and helped polishing the final manuscript.

## REFERENCES

- Al-Babili S, Bouwmeester HJ. 2015.** Strigolactones, a novel carotenoid-derived plant hormone. *Annual Review of Plant Biology* **66**: 161-186.
- Albacete A, Martinez-Andujar C, Martinez-Perez A, Thompson AJ, Dodd IC, Perez-Alfocea F. 2015.** Unravelling rootstock x scion interactions to improve food security. *Journal of Experimental Botany* **66**(8): 2211-2226.
- Bharti N, Tripathi S, Bhatla SC. 2015.** Photomodulation of strigolactone biosynthesis and accumulation during sunflower seedling growth. *Plant Signaling and Behavior* **10**(8): e1049792.
- Brewer PB, Yoneyama K, Filardo F, Meyers E, Scaffidi A, Frickey F, Akiyama K, Seto Y, Dun EA, Cremer JE, et al. 2016.** LATERAL BRANCHING OXIDOREDUCTASE acts in the final stages of strigolactone biosynthesis in Arabidopsis. *Proceedings of the National Academy of Sciences of the United States of America* **113**(22): 6301-6306.
- Cantero-Navarro E, Romero-Aranda R, Fernández-Muñoz R, Martínez-Andújar C, Pérez-Alfocea F, Albacete A. 2016.** Improving agronomic water use efficiency in tomato by rootstock-mediated hormonal regulation of leaf biomass. *Plant Science* **in press**. doi: [10.1016/j.plantsci.2016.03.001](https://doi.org/10.1016/j.plantsci.2016.03.001)
- Cheng X, Ruyter-Spira C, Bouwmeester H. 2013.** The interaction between strigolactones and other plant hormones in the regulation of plant development. *Frontiers in Plant Science* **4**: 199.
- Chevalier F, Nieminen K, Sanchez-Ferrero JC, Rodriguez ML, Chagoyen M, Hardtke CS, Cubas P. 2014.** Strigolactone promotes degradation of DWARF14, an alpha/beta hydrolase essential for strigolactone signaling in Arabidopsis. *Plant Cell* **26**(3): 1134-1150.
- Christmann A, Moes D, Himmelbach A, Yang Y, Tang Y, Grill E. 2006.** Integration of abscisic acid signalling into plant responses. *Plant Biology (Stuttgart, Germany)* **8**(3): 314-325.
- Christmann A, Weiler EW, Steudle E, Grill E. 2007.** A hydraulic signal in root-to-shoot signalling of water shortage. *Plant Journal* **52**(1): 167-174.
- Comstock JP. 2002.** Hydraulic and chemical signalling in the control of stomatal conductance and transpiration. *Journal of Experimental Botany* **53**(367): 195-200.
- Daszkowska-Golec A, Szarejko I. 2013.** Open or close the gate – stomata action under the control of phytohormones in drought stress conditions. *Frontiers in Plant Science* **4**: 138.
- Davies WJ, Zhang J. 1991.** Root signals and the regulation of growth and development of plants in drying soil. *Annual Review of Plant Physiology and Plant Molecular Biology* **42**: 55-76.

- Flokova K, Tarkowska D, Miersch O, Strnad M, Wasternack C, Novak O. 2014.** UHPLC-MS/MS based target profiling of stress-induced phytohormones. *Phytochemistry* **105**: 147-157.
- Foo E, Bullier E, Goussot M, Foucher F, Rameau C, Beveridge CA. 2005.** The branching gene *RAMOSUS1* mediates interactions among two novel signals and auxin in pea. *Plant Cell* **17**(2): 464-474.
- Foo E, Morris SE, Parmenter K, Young N, Wang H, Jones A, Rameau C, Turnbull CG, Beveridge CA. 2007.** Feedback regulation of xylem cytokinin content is conserved in pea and Arabidopsis. *Plant Physiology* **143**(3): 1418-1428.
- Foo E, Turnbull CG, Beveridge CA. 2001.** Long-distance signaling and the control of branching in the *rms1* mutant of pea. *Plant Physiology* **126**(1): 203-209.
- Gambino G, Perrone I, Gribaudo I. 2008.** A rapid and effective method for RNA extraction from different tissues of grapevine and other woody plants. *Phytochemical Analysis* **19**(6): 520-525.
- Ha CV, Leyva-Gonzalez MA, Osakabe Y, Tran UT, Nishiyama R, Watanabe Y, Tanaka M, Seki M, Yamaguchi S, Dong NV, et al. 2014.** Positive regulatory role of strigolactone in plant responses to drought and salt stress. *Proceedings of the National Academy of Sciences of the United States of America* **111**(2): 851-856.
- Hamiaux C, Drummond RS, Janssen BJ, Ledger SE, Cooney JM, Newcomb RD, Snowden KC. 2012.** DAD2 is an alpha/beta hydrolase likely to be involved in the perception of the plant branching hormone, strigolactone. *Current Biology* **22**(21): 2032-2036.
- Hartung W, Slovik S. 1991.** Physicochemical properties of plant growth regulators and plant tissues determine their distribution and redistribution: stomatal regulation by abscisic acid in leaves. *New Phytologist* **119**: 361-382.
- Holbrook NM, Shashidhar VR, James RA, Munns R. 2002.** Stomatal control in tomato with ABA-deficient roots: response of grafted plants to soil drying. *Journal of Experimental Botany* **53**(373): 1503-1514.
- Huber AE, Bauerle TL. 2016.** Long-distance plant signaling pathways in response to multiple stressors: the gap in knowledge. *Journal of Experimental Botany* **67**(7): 2063-2079.
- Johnson X, Breich T, Dun EA, Goussot M, Haurogne K, Beveridge CA, Rameau C. 2006.** Branching genes are conserved across species. Genes controlling a novel signal in pea are coregulated by other long-distance signals. *Plant Physiology* **142**(3): 1014-1026.
- Kohlen W, Charnikhova T, Lammers M, Pollina T, Toth P, Haider I, Pozo MJ, de Maagd RA, Ruyter-Spira C, Bouwmeester HJ, et al. 2012.** The tomato CAROTENOID CLEAVAGE DIOXYGENASE8 (SICCD8) regulates rhizosphere signaling, plant architecture and affects reproductive development through strigolactone biosynthesis. *New Phytologist* **196**(2): 535-547.
- Kohlen W, Charnikhova T, Liu Q, Bours R, Domagalska MA, Beguerie S, Verstappen F, Leyser O, Bouwmeester H, Ruyter-Spira C. 2011.** Strigolactones are transported through the xylem and play a key role in shoot architectural response to phosphate deficiency in nonarbuscular mycorrhizal host Arabidopsis. *Plant Physiology* **155**(2): 974-987.
- Kretschmar T, Kohlen W, Sasse J, Borghi L, Schlegel M, Bachelier JB, Reinhardt D, Bours R, Bouwmeester HJ, Martinoia E. 2012.** A petunia ABC protein controls strigolactone-dependent symbiotic signalling and branching. *Nature* **483**(7389): 341-344.
- Liu J, He H, Vitali M, Visentin I, Charnikhova T, Haider I, Schubert A, Ruyter-Spira C, Bouwmeester HJ, Lovisolo C, et al. 2015.** Osmotic stress represses strigolactone biosynthesis in *Lotus japonicus* roots: exploring the interaction between strigolactones and ABA under abiotic stress. *Planta* **241**(6): 1435-1451.
- Liu J, Novero M, Charnikhova T, Ferrandino A, Schubert A, Ruyter-Spira C, Bonfante P, Lovisolo C, Bouwmeester HJ, Cardinale F. 2013.** CAROTENOID CLEAVAGE DIOXYGENASE 7 modulates plant growth, reproduction, senescence, and determinate

nodulation in the model legume *Lotus japonicus*. *Journal of Experimental Botany* **64**(7): 1967-1981.

**Lopez-Raez JA, Kohlen W, Charnikhova T, Mulder P, Undas AK, Sergeant MJ, Verstappen F, Bugg TD, Thompson AJ, Ruyter-Spira C, et al. 2010.** Does abscisic acid affect strigolactone biosynthesis? *New Phytologist* **187**(2): 343-354.

**Manzi M, Lado J, Rodrigo MJ, Zacarias L, Arbona V, Gomez-Cadenas A. 2015.** Root ABA accumulation in long-term water-stressed plants is sustained by hormone transport from aerial organs. *Plant & Cell Physiology* **56**(12): 2457-2466.

**Matusova R, Rani K, Verstappen FW, Franssen MC, Beale MH, Bouwmeester HJ. 2005.** The strigolactone germination stimulants of the plant-parasitic *Striga* and *Orobanch* spp. are derived from the carotenoid pathway. *Plant Physiology* **139**(2): 920-934.

**Munoz-Espinoza VA, Lopez-Climent MF, Casaretto JA, Gomez-Cadenas A. 2015.** Water stress responses of tomato mutants impaired in hormone biosynthesis reveal abscisic acid, jasmonic acid and salicylic acid interactions. *Frontiers in Plant Science* **6**: 997.

**Ruiz-Lozano JM, Aroca R, Zamarreno AM, Molina S, Andreo-Jimenez B, Porcel R, Garcia-Mina JM, Ruyter-Spira C, Lopez-Raez JA. 2016.** Arbuscular mycorrhizal symbiosis induces strigolactone biosynthesis under drought and improves drought tolerance in lettuce and tomato. *Plant, Cell & Environment* **39**(2): 441-452.

**Ruyter-Spira C, Al-Babili S, van der Krol S, Bouwmeester H. 2013.** The biology of strigolactones. *Trends in Plant Science* **18**(2): 72-83.

**Sasse J, Simon S, Gubeli C, Liu GW, Cheng X, Friml J, Bouwmeester H, Martinoia E, Borghi L. 2015.** Asymmetric localizations of the ABC transporter PaPDR1 trace paths of directional strigolactone transport. *Current Biology* **25**(5): 647-655.

**Scholander PF, Bradstreet ED, Hemmingsen EA, Hammel HT. 1965.** Sap pressure in vascular plants: negative hydrostatic pressure can be measured in plants. *Science* **148**(3668): 339-346.

**Sorefan K, Booker J, Haurogne K, Goussot M, Bainbridge K, Foo E, Chatfield S, Ward S, Beveridge C, Rameau C, et al. 2003.** *MAX4* and *RMS1* are orthologous dioxygenase-like genes that regulate shoot branching in Arabidopsis and pea. *Genes & Development* **17**(12): 1469-1474.

**Torres-Vera R, Garcia JM, Pozo MJ, Lopez-Raez JA. 2014.** Do strigolactones contribute to plant defence? *Molecular Plant Pathology* **15**(2): 211-216.

**Tramontini S, Doering J, Vitali M, Ferrandino A, Stoll M, Lovisolo C. 2014.** Soil water-holding capacity mediates hydraulic and hormonal signals of near-isohydric and near-anisohydric *Vitis* cultivars in potted grapevines. *Functional Plant Biology* **41**(10-11): 1119-1128.

**Vogel JT, Walter MH, Giavalisco P, Lytovchenko A, Kohlen W, Charnikhova T, Simkin AJ, Goulet C, Strack D, Bouwmeester HJ, et al. 2010.** *SICC7* controls strigolactone biosynthesis, shoot branching and mycorrhiza-induced apocarotenoid formation in tomato. *Plant Journal* **61**(2): 300-311.

**Wilkinson S, Corlett JE, Oger L, Davies WJ. 1998.** Effects of xylem pH on transpiration from wild-type and *flacca* tomato leaves. A vital role for abscisic acid in preventing excessive water loss even from well-watered plants. *Plant Physiology* **117**(2): 703-709.

**Xiong L, Zhu JK. 2003.** Regulation of abscisic acid biosynthesis. *Plant Physiology* **133**(1): 29-36.

## SUPPORTING INFORMATION

**Figure S1. Relative soil water content (RWC) and water potential of soil ( $\Psi_{\text{soil}}$ ) during the course of a drought experiment**

**Figure S2. Effect of drought on SL amounts in tomato roots**

565 **Figure S3. Physiological performances of the grafted lines in the absence and presence of stress**  
566 **as a function of time**

567 **Figure S4. Transcript amounts of key SL biosynthetic genes as a function of leaf water potential**

568 **Figure S5. Effect of drought on free ABA as a function of time, and on transcript amounts of the**  
569 **ABA biosynthetic gene *SINCE1* as a function on leaf water potential**

570 **Table S1. List of primers**

571

## 572 **FIGURE LEGENDS**

573 **Figure 1. Physiological performances of the grafted lines in the absence and presence of stress.**  
574 Stomatal conductance **(a)**, and mean carbon assimilation rate **(b)** as a function on leaf water  
575 potential ( $\Psi_{\text{leaf}}$ ) of grafted tomato plants (WT/WT, SL-/SL- and WT/SL-) along a water-deprivation  
576 time-course. Full symbols in each series indicate rehydrated samples (recovery). Data represent  
577 the mean and SEM of  $n = 6$  biological replicates from 2 independent experiments.

578

579 **Figure 2. Effect of drought on the transcript amounts of SL biosynthetic genes (*SICCD7* and**  
580 ***SICCD8*) of roots (a-b) and shoots (c-d) of grafted tomato plants (WT/WT, SL-/SL- and WT/SL-)**  
581 **during a time-course (0, 1, 3 and 5 days from water withdrawal for T0 through T5). R indicates the**  
582 **rehydrated samples (recovery). Gene transcript abundance was normalized to endogenous *EF1α***  
583 **and presented as fold-change value over WT/WT at T0, which was set to 1. Data represent the**  
584 **mean and SEM of  $n = 6$  biological replicates from 2 independent experiments. Different letters**  
585 **indicate significant differences between plant lines for the same time point, as determined by a**  
586 **two-way ANOVA test ( $P < 0.05$ ). n.d. = not detectable.**

587

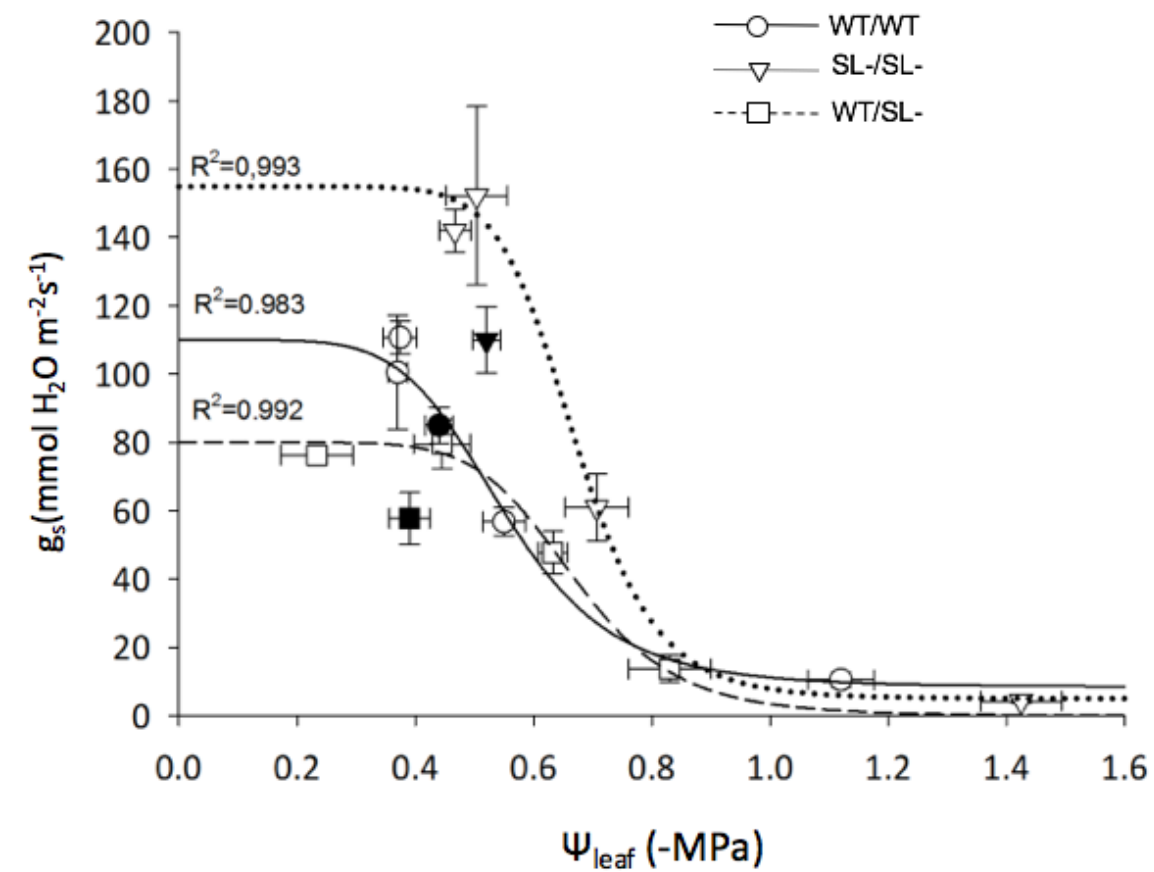
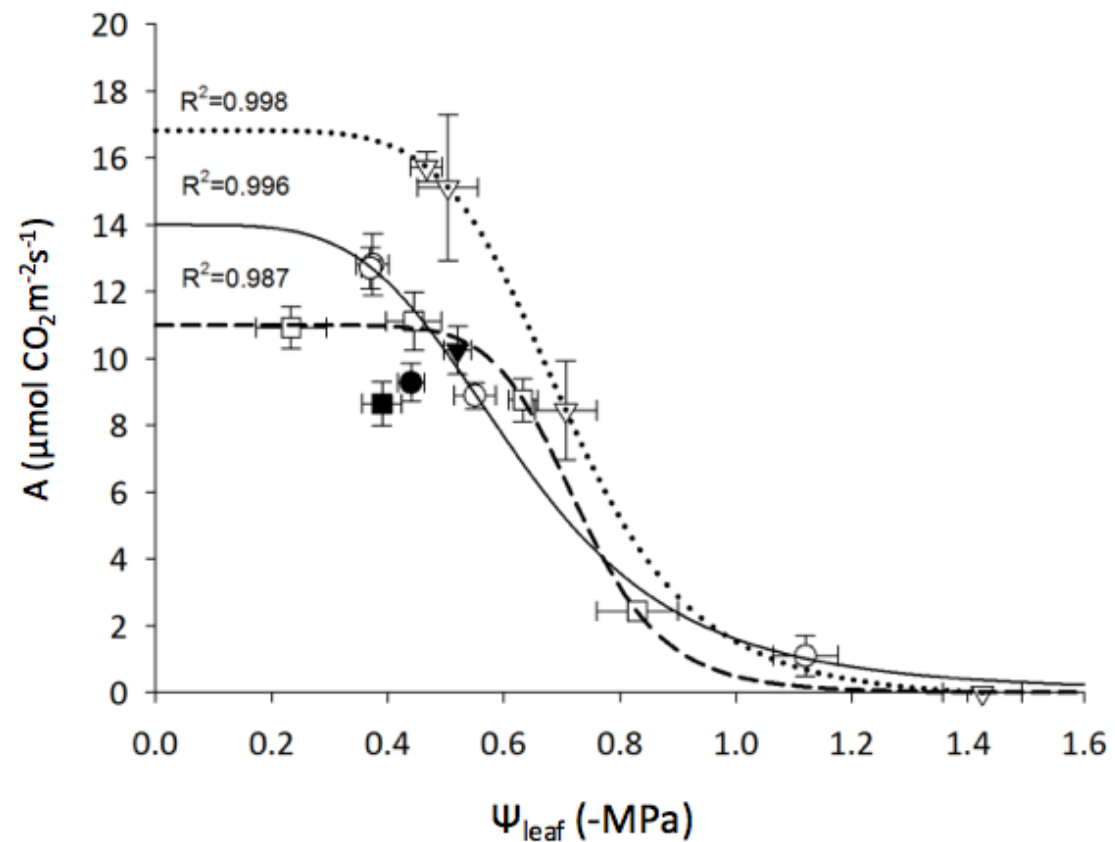
588 **Figure 3. Effect of drought on free ABA as a function on leaf water potential ( $\Psi_{\text{leaf}}$ ) and on**  
589 **transcript amounts of the ABA biosynthetic gene *SINCE1* in roots (a-b) and shoots (c-d) of**  
590 **grafted tomato plants (WT/WT, SL-/SL- and WT/SL-) during a time-course (0, 1, 3 and 5 days from**  
591 **water withdrawal for T0 through T5). Full symbols (a, c) or R (b, d) indicate the rehydrated samples**  
592 **(recovery). Gene transcript abundance was normalized to endogenous *EF1α* and presented as**  
593 **fold-change value over WT/WT at T0, which was set to 1. Data on ABA represent the mean and**  
594 **SEM of  $n = 4$  biological replicates (each replicate a pool of 2 plants) from 2 independent**  
595 **experiments. Data on *SINCE1* represent the mean and SEM of  $n = 6$  biological replicates from 2**

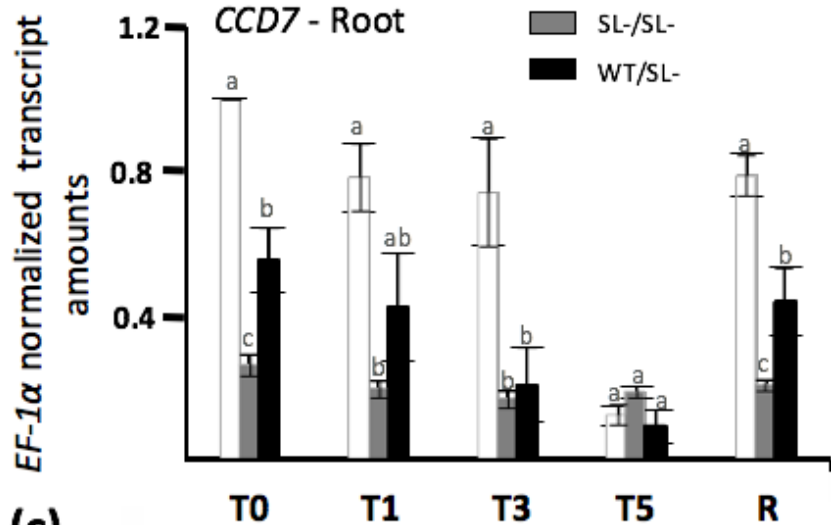
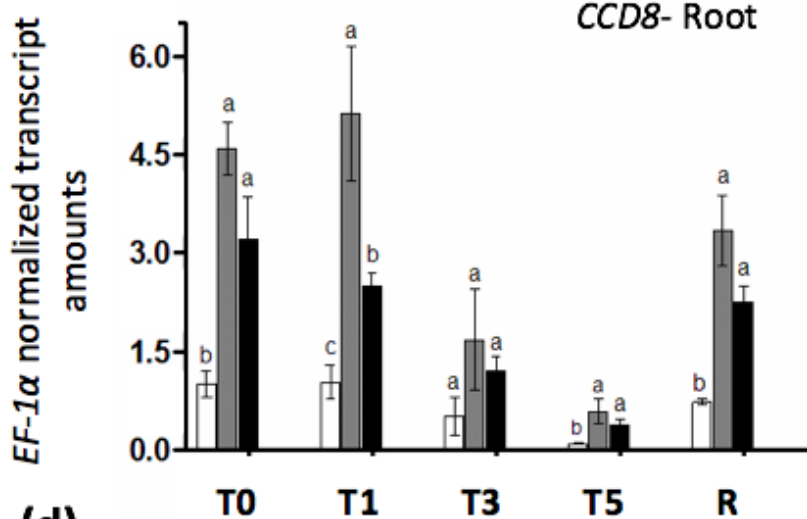
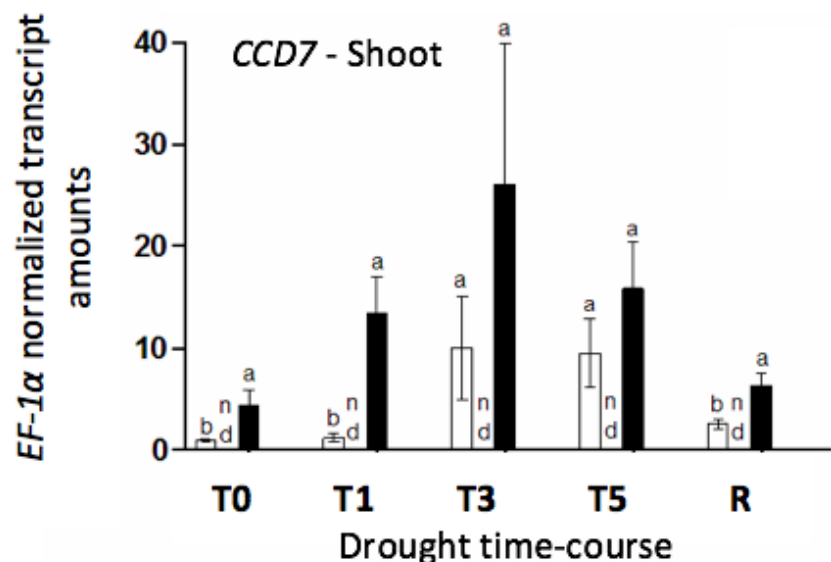
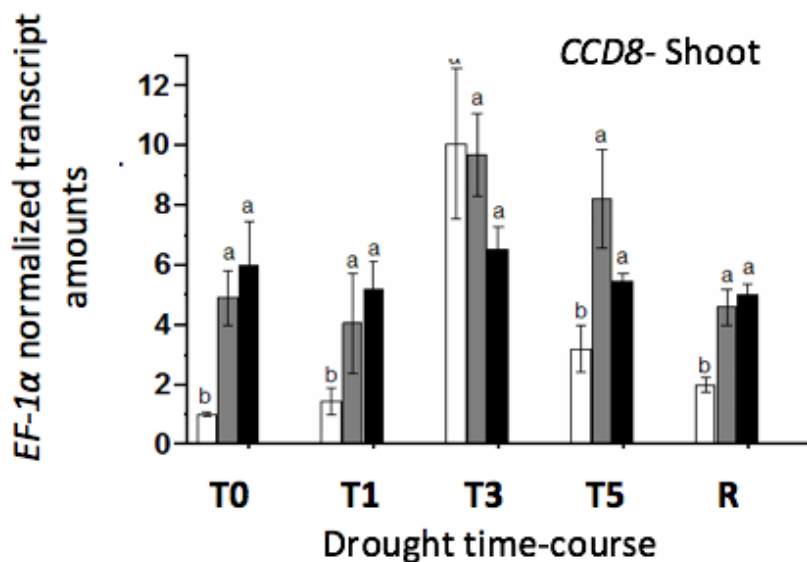
596 independent experiments. Different letters in **(b)** and **(d)** indicate significant differences between  
597 plant lines for the same time point, as determined by a two-way ANOVA test ( $P < 0.05$ ).  
598

599 **Figure 4. Dose-response of leaves to treatment with exogenous ABA** at different concentrations.  
600 Stomatal conductance was measured at 30-s intervals before and during ABA treatments  
601 performed on detached composite leaves from grafted tomato plants (WT/WT, SL-/SL-, WT/SL-).  
602 WT/WT plant pre-treated with 5  $\mu$ M *racGR24* were analysed only for the 5  $\mu$ M ABA treatment  
603 (black bar). Values represent the mean and SEM of at least  $n = 6$  biological replicates from 2  
604 independent experiments, and refer to the time (seconds) needed for the decrease of stomatal  
605 conductance to start, from the time of ABA addition to the dipping solution. Different letters  
606 indicate significant differences between plant lines for the same treatment, as determined by a  
607 two-way ANOVA test ( $P < 0.05$ ).  
608

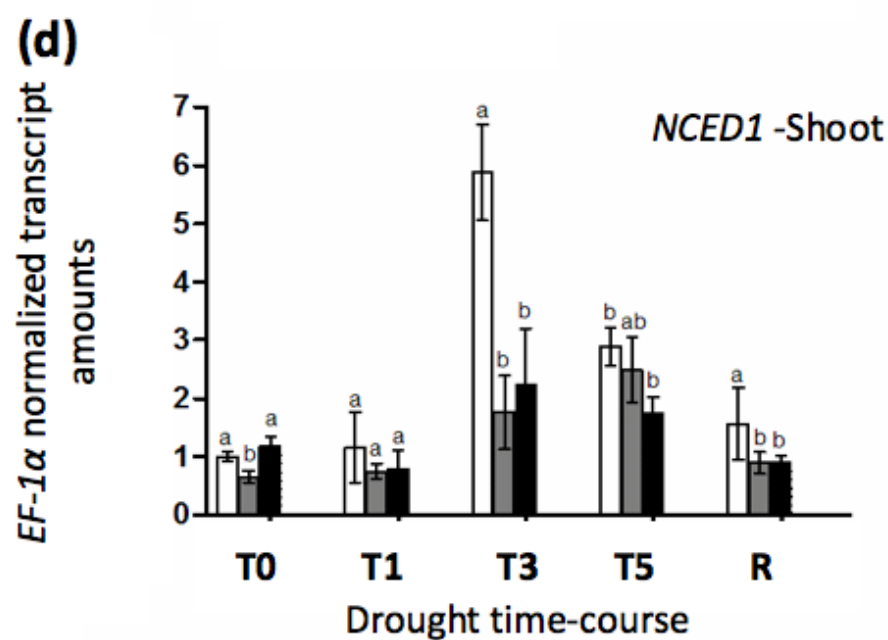
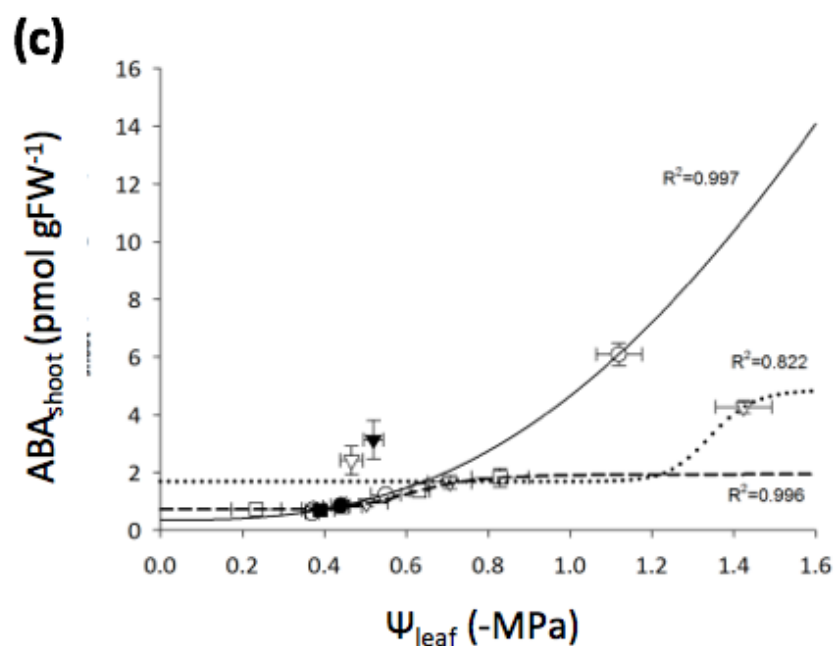
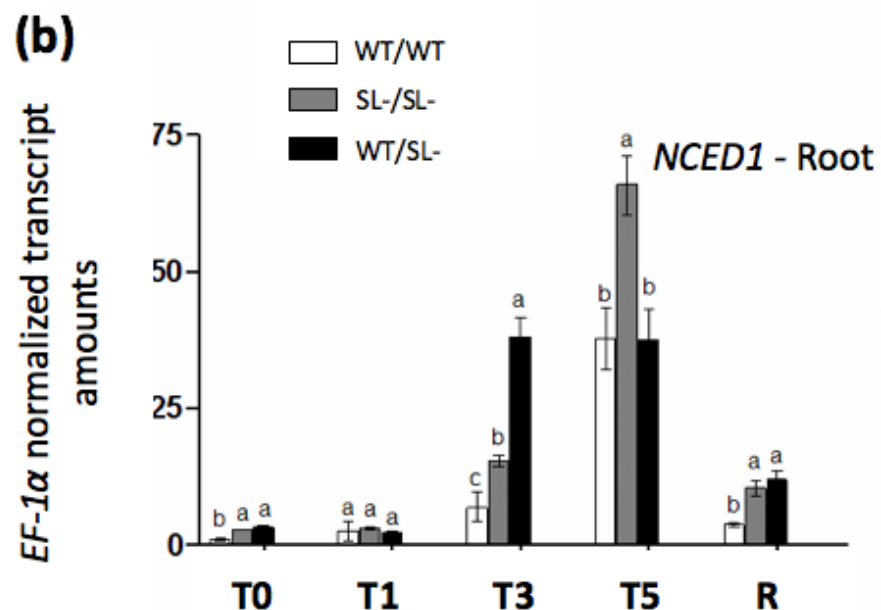
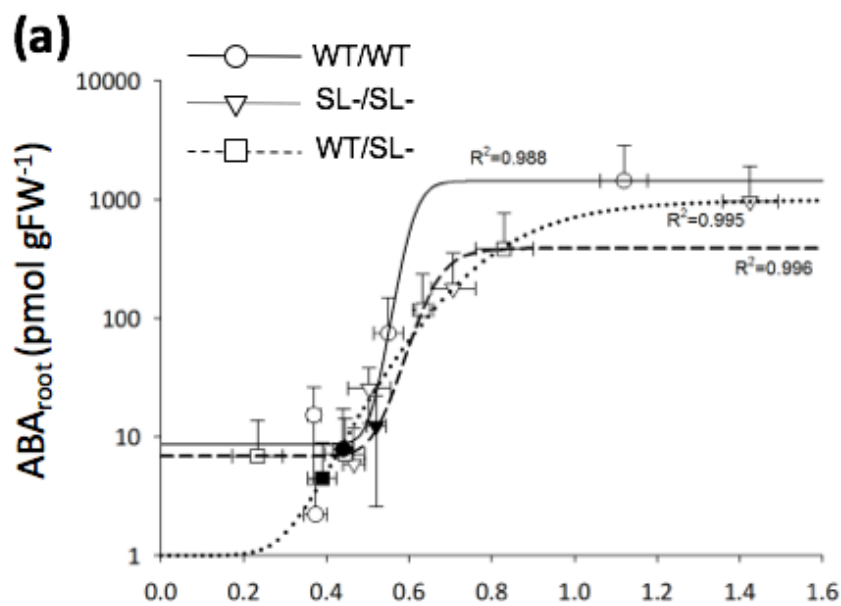
609 **Figure 5. Schematic drawing of the main connections between SL and ABA in roots and shoots of**  
610 **tomato under drought stress.** In the model, the effects of SL on ABA levels may be negative in the  
611 roots, as proven by *racGR24* treatment in *L. japonicus* (Liu *et al.*, 2015). Thereby, the drop in SL  
612 synthesis in this organ under osmotic (PEG-infused) stress may be needed but not necessarily  
613 sufficient to let ABA levels rise (results untested in other plant species so far; **1**). SL synthesis is  
614 inhibited in roots under osmotic/drought stress, so shootward SL flow decreases (**2**); in tomato,  
615 root-produced ABA is not translocated nor needed for appropriate shoot responses to stress  
616 (Holbrook *et al.*, 2002). The effects of shoot-produced or exogenous SL on ABA sensitivity of  
617 stomata are in turn positive (**3**) [(Ha *et al.*, 2014; Liu *et al.*, 2015) and this work]. SL flowing  
618 shootward inhibit the transcription of SL biosynthetic genes (thicker line, **4**), as reduced quantities  
619 in the upstream flow (or possibly, a second messenger – different than ABA - produced in the  
620 roots in response to low SL) are sufficient to let transcripts of SL biosynthetic genes increase  
621 (thinner line; **5**) and as a likely consequence, also sensitivity to ABA (**6**). Whether osmotic/drought  
622 stress can increase SL gene transcription and ABA sensitivity in the shoots even if SL synthesis in  
623 the root is not decreased is not known (question mark). Although SL remain undetectable in  
624 whole-shoot analyses of stressed tomato, localized accumulation may occur, as proposed (Liu *et*  
625 *al.*, 2015) and suggested by transcript quantification of biosynthetic genes (Ha *et al.*, 2015; this  
626 work). Alternatively, steady-state SL levels may be needed and sufficient to ensure wild-type

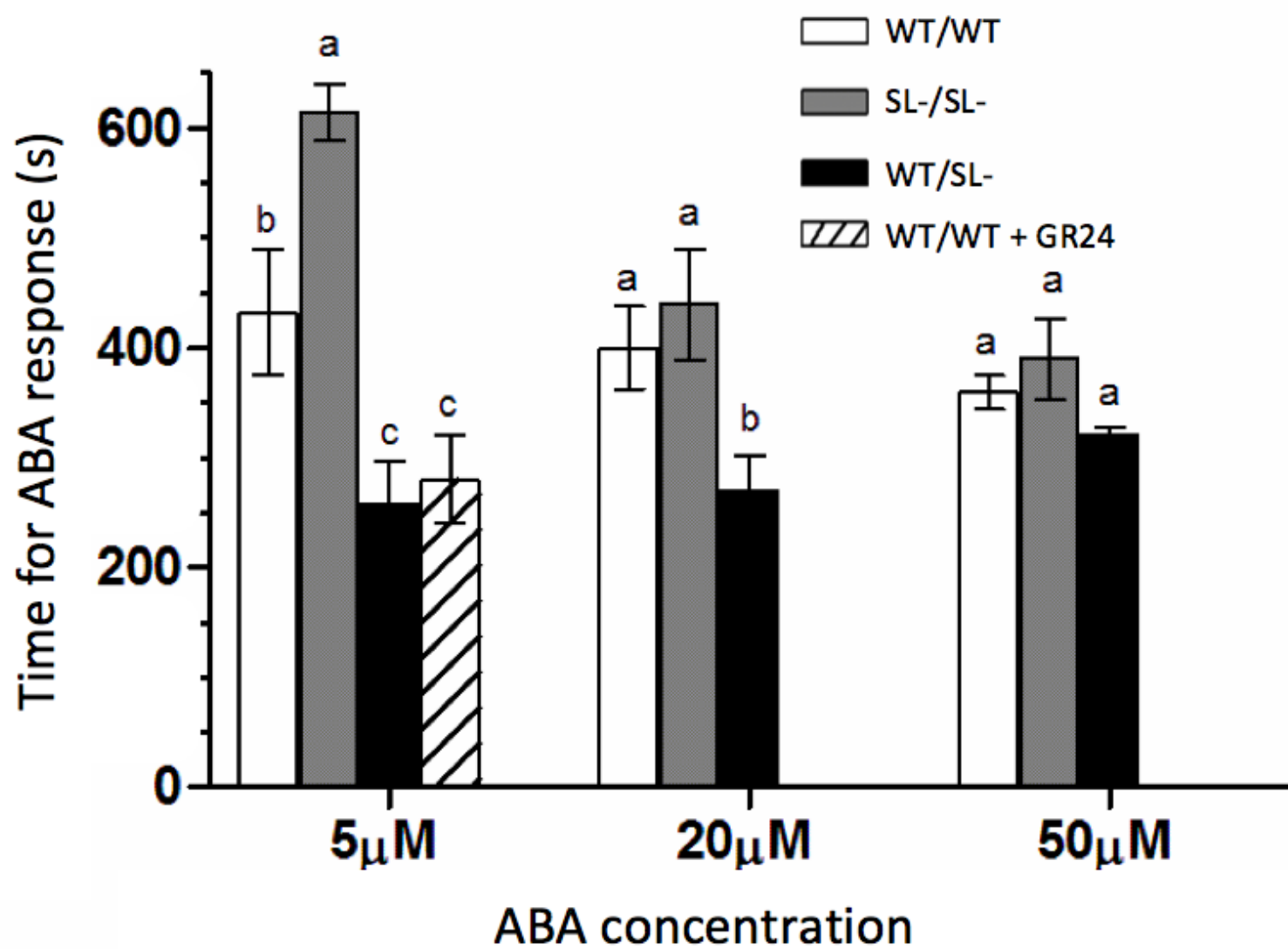
627 sensitivity to ABA in stressed shoot tissues; or other, yet unidentified SL(-like) molecules may be  
628 induced.  
629

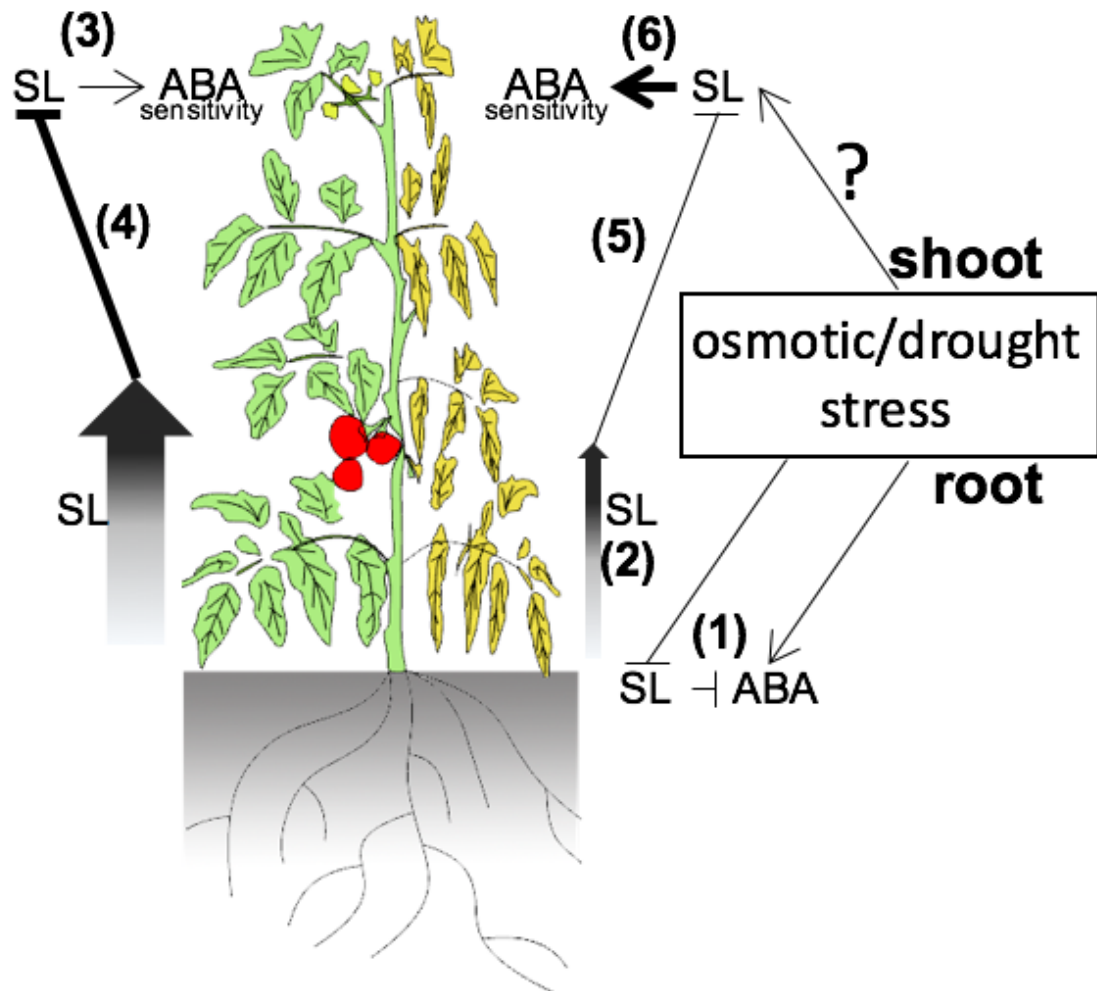
**(a)****(b)**

**(a)****(b)****(c)****(d)**









## **New Phytologist Supporting Information**

Article title: **Low levels of strigolactones in roots as a component of the systemic signal of drought stress in tomato**

Authors: Ivan Visentin, Marco Vitali, Manuela Ferrero, Yanxia Zhang, Carolien Ruyter-Spira, Ondřej Novák, Miroslav Strnad, Claudio Lovisolo, Andrea Schubert, Francesca Cardinale

Article acceptance date: 4 August 2016

The following Supporting Information is available for this article:

**Fig. S1 Relative soil water content (RWC) and water potential of soil ( $\Psi_{\text{soil}}$ ) during the course of a drought experiment**

**Fig. S2 Effect of drought on SL amounts in tomato roots**

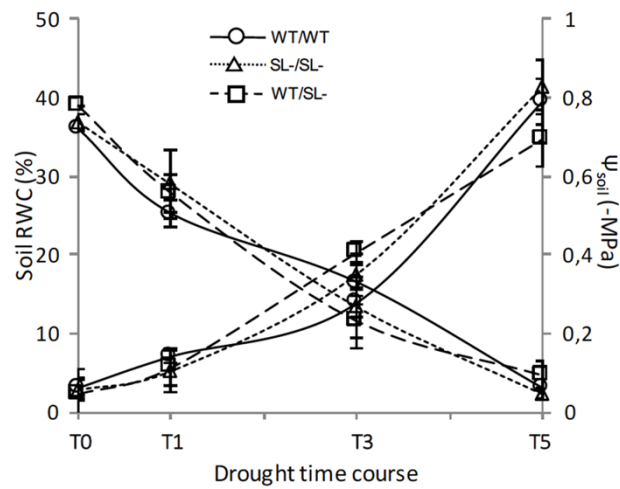
**Fig. S3 Physiological performances of the grafted lines in the absence and presence of stress as a function of time**

**Fig. S4. Transcript amounts of key SL biosynthetic genes as a function of leaf water potential**

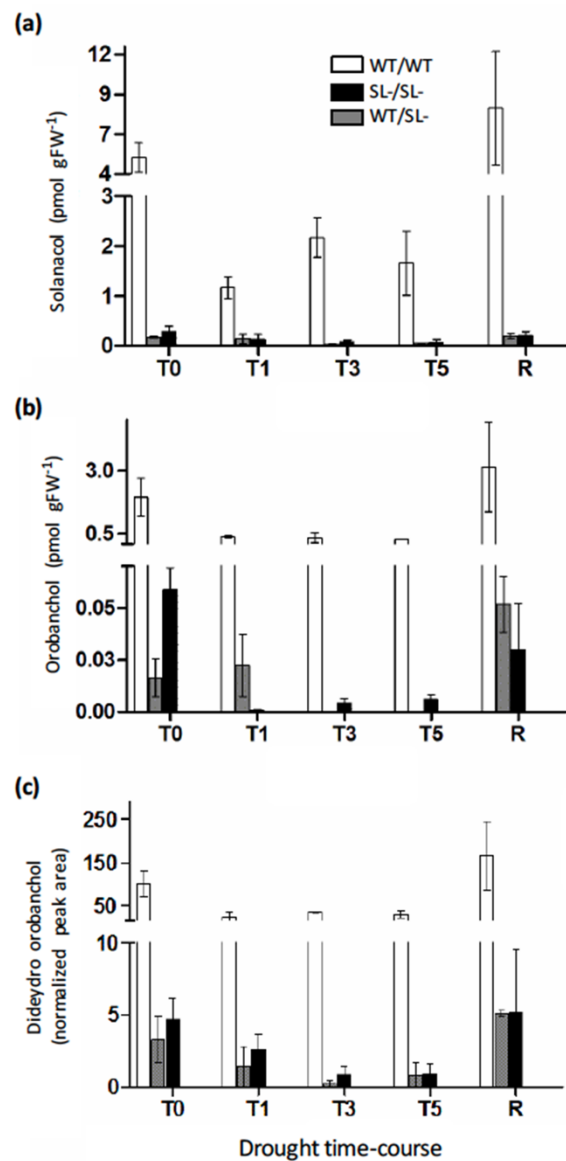
**Fig. S5 Effect of drought on free ABA as a function of time, and on transcript amounts of the ABA biosynthetic gene *SINCE1* as a function on leaf water potential**

**Table S1 List of primers**

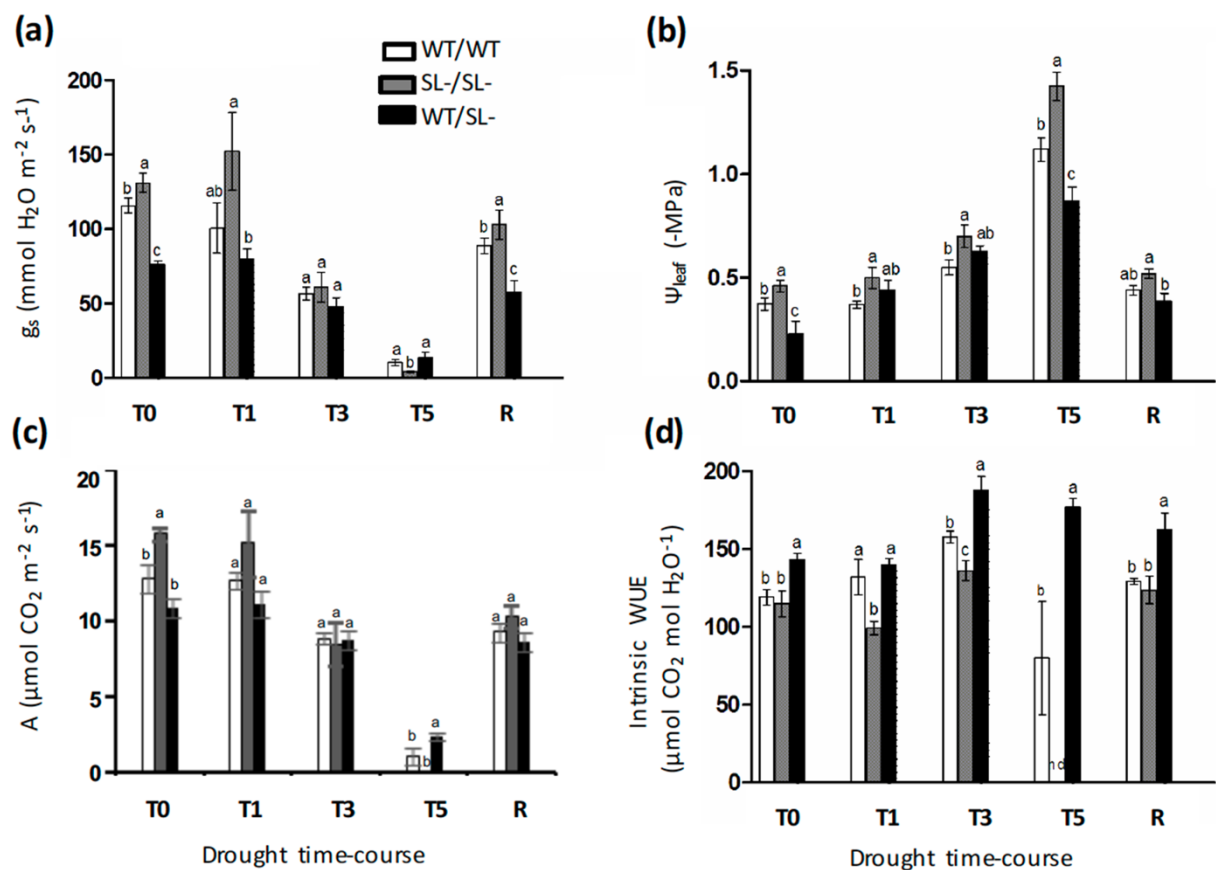
**Fig. S1 Relative water content (RWC) and water potential of soil ( $\Psi_{\text{soil}}$ ) during the course of a drought experiment.** Soil RWC was gravimetrically determined by collecting daily soil from three different points and depths in each pot, to assess water content after oven drying. At the same time, the soil water retention curve was assessed with pressure plate measurements of the potting substrate (Tramontini S, Doering J, Vitali M, Ferrandino A, Stoll M, Lovisolo C. 2014. Soil water-holding capacity mediates hydraulic and hormonal signals of near-isohydric and near-anisohydric *Vitis* cultivars in potted grapevines. *Functional Plant Biology* 41(10-11): 1119-1128). For both datasets values represent the mean and SEM of  $n = 6$  samples from two independent experiments.



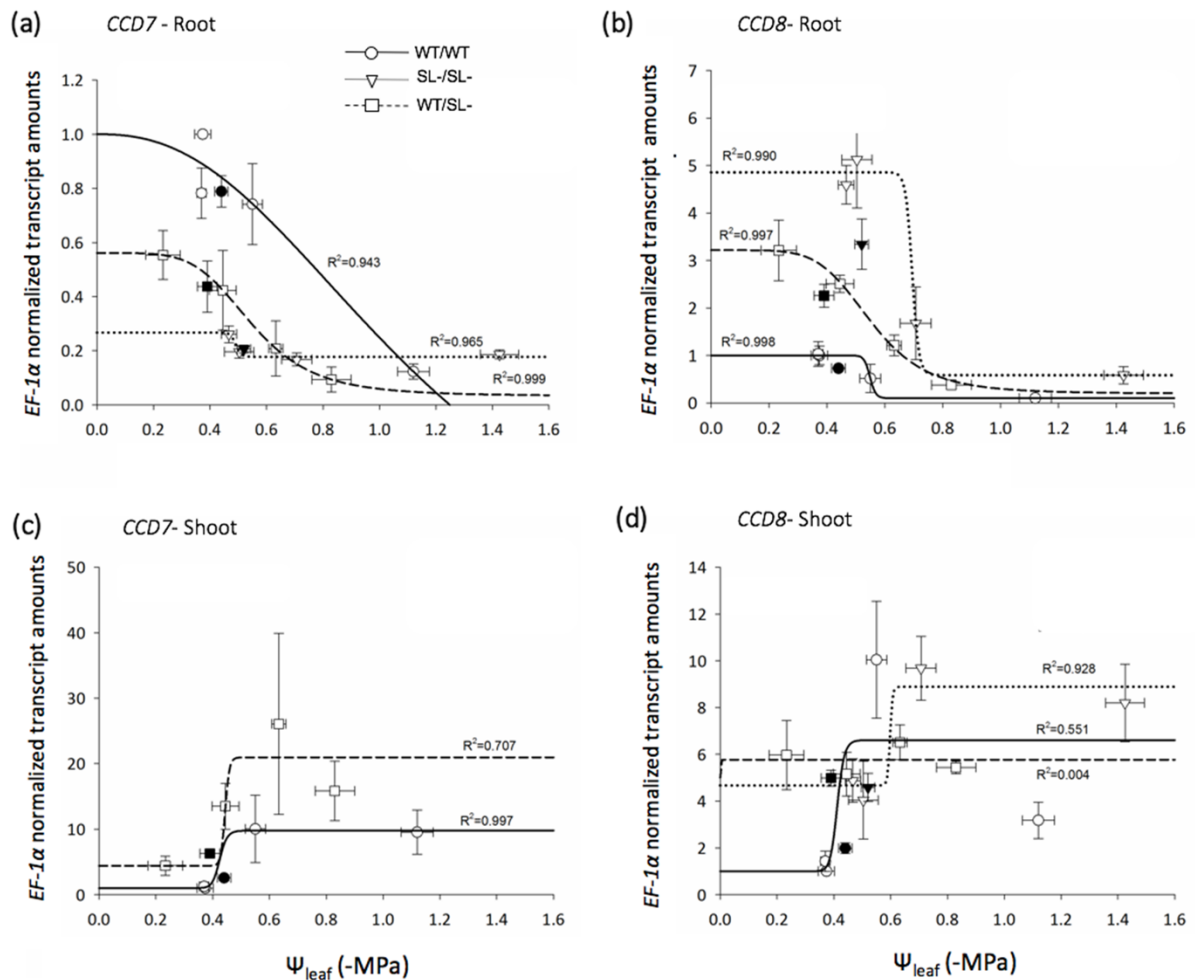
**Fig. S2 Effect of drought on SL biosynthesis in tomato roots:** solanacol (a), orobanchol (b) and the didehydro-orobanchol isomer with retention time 4'6'' (c) were quantified in roots of grafted plants (WT/WT, SL-/SL- and WT/SL-) along a time-course (0, 1, 3 and 5 days from the beginning of stress for T0 through T5). R indicates the rehydrated (Recovery) samples. Data represent the mean and SEM of  $n = 2$  samples derived from the pool of 3 plants each, in two independent experiments. While solanacol and orobanchol are expressed as absolute amounts per g of fresh tissue weight, the didehydro isomer of orobanchol is expressed as a percentage ratio of MS/MS peak area normalized over values for WT tissues at T0, due to the lack of a standard.



**Fig. S3. Physiological performances of the grafted lines in the absence and presence of stress as a function of time.** Stomatal conductance (a), leaf water potential (b) and mean carbon assimilation rate (c), and intrinsic water use efficiency (WUE) (d) of grafted tomato plants (WT/WT, SL-/SL- and WT/SL-) along a water-deprivation time-course (0, 1, 3 and 5 days from the beginning of stress for T0 through T5). R indicates rehydrated samples (recovery). Data represent the mean and SEM of  $n = 6$  biological replicates from 2 independent experiments. Different letters indicate significant differences within the same time point as determined by a two-way ANOVA test ( $P < 0.05$ ). n.d. = not detectable.

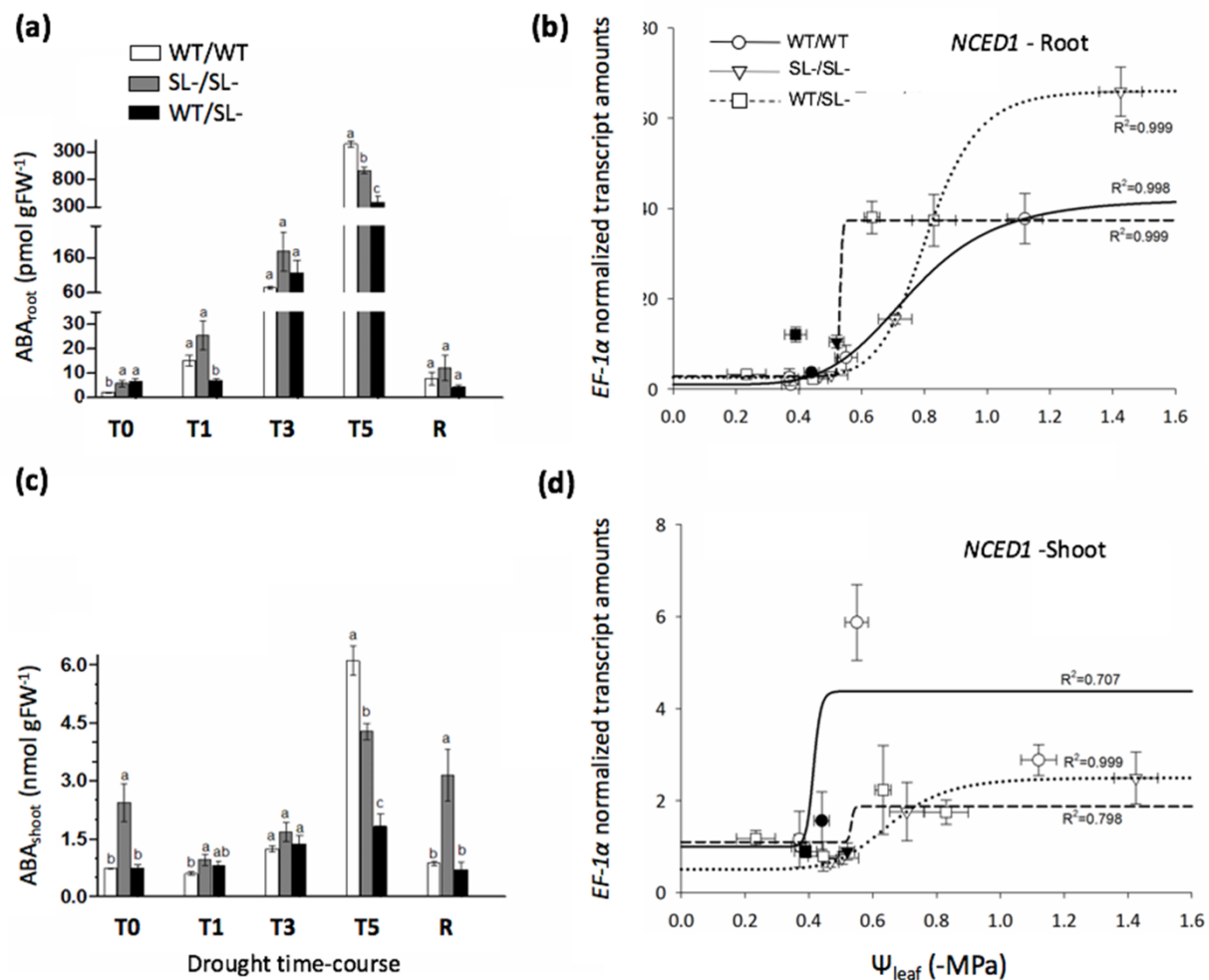


**Fig. S4. Transcript amounts of key SL biosynthetic genes (*SICCD7* and *SICCD8*) in roots (a-b) and shoots (c-d) of grafted tomato plants (WT/WT, SL-/SL- and WT/SL-) as a function of leaf water potential during a drought time-course (0, 1, 3 and 5 days from water withdrawal). Full symbols in each series indicate the rehydrated samples (recovery). Gene transcript abundance was normalized to endogenous *EF1 $\alpha$*  and presented as fold-change value over WT/WT at T0, which was set to 1. Data represent the mean and SEM of  $n = 6$  biological replicates from 2 independent experiments. *SICCD7* transcripts were undetectable in silenced (SL-) shoots.**





**Fig. S5. Effect of drought on free ABA and on transcript amounts of the ABA biosynthetic gene *SINCED1* as a function on leaf water potential in roots (a-b) and shoots (c-d) of grafted tomato plants (WT/WT, SL-/SL- and WT/SL-) during a time-course (0, 1, 3 and 5 days from water withdrawal for T0 through T5). Full symbols (a, c) or R (b, d) indicate the rehydrated samples (recovery). Gene transcript abundance was normalized to endogenous *EF1α* and presented as fold-change value over WT/WT at T0, which was set to 1. Data on ABA represent the mean and SEM of  $n = 4$  biological replicates (each replicate a pool of 2 plants) from 2 independent experiments. Data on *SINCED1* represent the mean and SEM of  $n = 6$  biological replicates from 2 independent experiments. Different letters in (a) and (c) indicate significant differences between plant lines for the same time point, as determined by a two-way ANOVA test ( $P < 0.05$ ).**



**Table S1** Primer pairs for transcript quantification via qRT-PCR.

**Kohlen W, Charnikhova T, Lammers M, Pollina T, Toth P, Haider I, Pozo MJ, de Maagd RA, Ruyter-Spira C, Bouwmeester HJ, et al. 2012.** The tomato CAROTENOID CLEAVAGE DIOXYGENASE8 (SLCCD8) regulates rhizosphere signaling, plant architecture and affects reproductive development through strigolactone biosynthesis. *New Phytologist* **196**(2): 535-547.

**Lopez-Raez JA, Kohlen W, Charnikhova T, Mulder P, Undas AK, Sergeant MJ, Verstappen F, Bugg TD, Thompson AJ, Ruyter-Spira C, et al. 2010.** Does abscisic acid affect strigolactone biosynthesis? *New Phytologist* **187**(2): 343-354.

Gene ID	Forward primer	Reverse primer	Reference
<i>SLCCD7</i>	GTTGCTCTTACCAATGGTTCAATTT	TACATTCATCATGGAAGGATCAAAGTT	(Kohlen <i>et al.</i> , 2012)
<i>SLCCD8</i>	CCAATTGCCTGTAATAGTTCC	GCCTTCAACGACGAGTTCTC	(Kohlen <i>et al.</i> , 2012)
<i>SINCE1</i>	ACCCACGAGTCCAGATTTC	GGTTCAAAAAGAGGGTTAG	(Lopez-Raez <i>et al.</i> , 2010)
<i>SIEF1<math>\alpha</math></i>	GATTGGTGGTATTGGAAGTGC	AGCTTCGTGGTGCATCTC	(Kohlen <i>et al.</i> , 2012)

**Supplementary Table S1**

Gene ID	Forward primer	Reverse primer	Reference
<i>SLCCD7</i>	GTTGCTCTTACCAATGGTTCAATTT	TACATTCATCATGGAAGGATCAAAGTT	(Kohlen <i>et al.</i> , 2012)
<i>SICCD8</i>	CCAATTGCCTGTAATAGTTCC	GCCTTCAACGACGAGTTCTC	(Kohlen <i>et al.</i> , 2012)
<i>SINCED1</i>	ACCCACGAGTCCAGATTTC	GGTTCAAAAAGAGGGTTAG	(Lopez-Raez <i>et al.</i> , 2010)
<i>SIEF1<math>\alpha</math></i>	GATTGGTGGTATTGGAAGTCTC	AGCTTCGTGGTGCATCTC	(Kohlen <i>et al.</i> , 2012)

Primer pairs for transcript quantification via qRT-PCR